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# Effects of Collector Sea Urchin (*Tripneustes gratilla*) on the Reduction of Brown Seaweed (*Dictyota* spp. and *Padina* spp.) Cover in Post-Coral Bleached Systems

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**Abstract** – The purpose of this research is to determine the effects of *Tripneustes gratilla* in reducing the abundance of *Dictyota dichotoma* and *Padina* spp. in macroalgal-dominated marine ecosystems. Corals attached with macroalgae were exposed to the *Tripneustes gratilla* for two weeks. Measurement of the macroalgal density before and after the exposure of *T. gratilla* was administered. The Fleshy Macroalgal Index of the units with sea urchin and without sea urchin were also compared and analyzed. A software, PhotoQuad, was utilized to quantify the FMI of each unit. The results showed that the presence of *T. gratilla* did not significantly reduce ( $p=0.23$ ) the macroalgal cover in comparison to the control group. Nevertheless, the compared FMI of the pre-exposure and post-exposure to the urchin showed a significant difference ( $p=0.023$ ). In conclusion, the reduction of the macroalgal cover may be attributed to the presence of *Tripneustes gratilla*. Although it is only comparably different, the exposure to *Tripneustes gratilla* can potentially aid in the recovery of macroalgal-covered post coral bleached systems and in the control of invasive macroalgae population.

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**Introduction.** – Coral reefs are known as the most biologically diverse and productive ecosystem on the planet [1]. Massive bleaching of corals has been observed over the past two decades in different parts of the world's ocean [?]; Shenkar et al. 2005) and considered as the most serious threats for coral reefs [2]. It is caused by the disruption of the symbiotic interaction between the coral host and the symbiotic algae, Symbiodinium inducing a massive expulsion of these symbionts or loss of their pigments occur [2, 3, 14]. This phenomenon can be linked to global climate change and increasing ocean temperatures [4].

Coral mortality following bleaching events provides space on the reef for rapid colonization by turf-forming algae [5], which compete with corals for space, and prevent coral recover (Tanner 1995). Macroalgae produces allelochemicals which can influence the growth, survival and reproduction of other organisms within their vicinity [6]. They are known to suppress the settlement and fitness of corals which is why coral reefs that are dominated by macroalgae can lead to further decline [15] Haan J 2015). Results from previous studies showed that the coral-algal

interaction have significant, species-specific, negative effects on the growth and survival of corals [7]. The dominant scleractinian group, *Acropora* spp., is found to be the most strongly impacted by contact with allelopathic macroalgae [15]. It is also highlighted that the outcome of the macroalgal-coral competition can have significant implications on the growth of corals [7].

The presence of sea urchins can influence the community structure of habitats containing marine plants and algae populations [8]. They are usually found in areas abundant in algae. The collector sea urchin (*Tripneustes gratilla*) is an herbivore widely found in tropical and subtropical areas [9]. Their abundance and distribution in coral reef areas suggest that they play an important role ecologically and is vital to the flow of matter and energy [10].

**Methods.** – This chapter discusses the methods used in conducting the study and in collecting the data. The research experiment is divided into four phases: collection of the samples needed, exposure of the macroalgae to

the coral fragments, exposure of the sea urchin to the macroalgal-coral setup and comparison of initial and final data of control and experimental setup.

*Experimental Set-up.* The experimental set-up of this study was based on the study of Rasher and Hay (2011) entitled Chemically rich seaweeds poison corals when not controlled by herbivores. The set-up was established in Piagau Island, Brgy. La Paz, Nueva Valencia, Guimaras, Philippines. The metal cage has a total of ten units. Five units of coral and macroalgae were prepared in the cage for the control group. As for the treatment group, five units of coral, macroalgae, and sea urchin were prepared. Randomized Complete Block Design (RCBD) was used as the experimental design. In RCBD, replicates are separated randomly and each treatment has an equal probability of being assigned to a given experimental unit.

*Collection of Samples.* The researchers, with the assistance of professional divers, gathered bleached coral fragments of *Acropora spp.* found in Taklong Island National Marine Reserve, Guimaras, Philippines. These fragments are approximately the same length, five to six cm, for the standardization of the macroalgae-coral contact. It was collected using rubber gloves to avoid direct contact with the corals. The macroalgae were collected in Brgy. Lapaz, Nueva Valencia Guimaras having the same weight of five grams per unit. Lastly, the *Tripneustes gratilla* were collected in Brgy. Canhawan, Nueva Valencia, Guimaras with the assistance of authorized personnel. The researchers were assisted by experts in the field of identifying the organisms stated above to validate the identification of the organisms used in the study.

*Coral Planting.* The branches were glued individually into cement cones with a diameter of 8 cm using an underwater epoxy. Marine epoxy was used because there are no recorded negative effects as coral adhesives and it is effective in attachment and transplant survival (Dizon et al. 2008). It is essential to plant the corals in order to simulate the natural conditions of the macroalgal-coral competition. See fig. 1

*Placement of Corals in the Caged Racks.* The treatments and controls replicates (n=15 coral branches each) have separate metal racks. The metal racks were covered with 1x1 cm grid metal screen to avoid other herbivores from entering. The racks were placed on designated areas around the island. Each of the coral branches within the cement cones were interspersed 15-cm apart across the metal rack. As a means to minimize the extraneous variables and for easier measurement, there is a divider between each of the coral branches. The measurement of the whole metal rack is 115x46x50 cm (LxWxH). Three metal racks were used so that there would be 15 organisms for each experimental group for the statistical analysis.



Fig. 1: Coral branch planted into a cement cone

*Exposure of *D. dichotoma* and *Padina spp.* to *Acropora spp.** The macroalgae species *D. dichotoma* and *Padina spp.* were collected around the site. As seen in Figure 2 (appendix b), nails were embedded on the opposite side of each cement cone in order for the rope attached with the *D. dichotoma* and *Padina spp.* to be positioned over each nail head. In this way, the *D. dichotoma* and *Padina spp.* were in stable contact with the coral. The macroalgae used in the coral-macroalgal contact are found in the site. In order to avoid stress compounds that might be emitted from the deployed *D. dichotoma* and *Padina spp.*, the whole thalli was used. The coral branches were exposed to the macroalgal species for 14 days. Initial data will be collected before the exposure of *T. gratilla* to the treatments.

*Exposure of *T. gratilla* to the treatments.* The *T. gratilla* were collected in Brgy. Canhawan, Nueva Valencia, Guimaras. Only one *T. gratilla* will be exposed per unit. The exposure will last for 14 days.

*Measurement of the Fleshy Macroalgal Index (FMI).* The FMI will be measured using the photoquad method. Photoquad method provides visual estimation but in a digital version. This method is usually used in coral monitoring programs in field surveys [?]. The remaining macroalgae attached to the rope were photographed. After taking a picture, the photo quadrat analysis software, photoQuad, quantified the FMI using the grid cell count option. A direct estimate of the species cover was automatically performed. The software offers a more versatile, quick and accurate result. The FMI of each unit will be measured before and after the exposure *T. gratilla* for comparison.

*Data Analysis.* In order to compare results, statistical software PAST will be used. The statistical test T-test will be used in order to compare the means of FMI of each unit. The initial and final FMI of the controls were also compared and analyzed. The statistical software used in computing the paired sample test was IBM SPSS

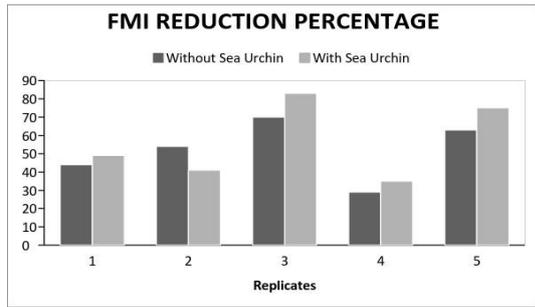


Fig. 2: Comparison of the Fleshy Macroalgal Index reduction values between units with and without sea urchin

Statistics 22.

*Proper Disposal* *Tripneustes gratilla* were returned to its natural habitat, *Acropora spp.* were replanted around the area of Piagau Island, and the remaining *Dictyota dichotoma* and *Padina spp* were disposed. The metal cages were left in their original locations to serve as a fish nursery, as prescribed by the Department of Environment and Natural Resources.

**Results and Discussion.** – There is a general reduction in percent coverage in all the experimental groups. Despite having a preference in consuming *Padina sanctae-crucis* compared to any other macroalgal species (Stimson et al. 2007). The presence of *T. gratilla* in post-coral bleached systems did not significantly reduce the macroalgal cover in comparison to the control group (See fig.2, refer to table 1). Although, it can be observed that the FMI reduction percentage of the treatments, in general, were greater compared to that of the controls. There was only a comparable difference. Nevertheless, the compared FMI of the pre- and post-exposure to the urchin showed a significant difference (See fig.3, refer to Table 2). This suggests that certain factors have probably affected the reduction of macroalgal cover of both experimental groups which has led to some discrepancies in the data acquired.

First probable factor may be the failure to control all possible existence of extraneous variables, such as the presence of other organism inside the cages. The unwanted organisms found in the cage were school of juvenile reef fishes and sea cucumbers. The cages were intentionally designed to restrict the entrance of these organisms, yet their natural biomechanisms has allowed their entrance. The presence of these organisms was beyond the control of the researchers. The small size of juvenile reef fishes and the capability of sea cucumbers (*Synapta maculata*) to congest themselves has enabled them to penetrate the cages (Elder Trueman, 1980). It is possible that these organisms have consumed the macroalgae in each unit, since these organisms are known herbivores (Hay, 1997; Martinez-Diaz and Perez-Espana, 1999). Second probable factor is the unstable weather

Table 1: Computed t-value between units with and without sea urchin based on the FMI

Fleshy Macroalgal Index (FMI)	
Variable	t-value
Cell count	0.3625
Area(cm <sup>2</sup> )	0.41675
Coverage percentage	0.23299

conditions. In a study conducted by Sinnot (2009), it concludes that water quality has a significant effect on the growth of macroalgae at certain levels. It has been shown that turbidity severely limits the capacity of macroalgae to photosynthesize and grow (Hay 1981). The varying parameters due to the unstable weather conditions during the data gathering might have greatly affected the macroalgal growth. Inconsistent environmental conditions could lower the metabolic rate [10] Third, the weight of the sea urchin might also have affected the percentage reduced because of the varying capacity of consumption. Because of the variance in the weight and sizes of the sea urchin, the rate of consumption also differs. The smaller the size of the sea urchin corresponds to a higher ingestion rate of the organism [10]. Lastly, it may also be caused the deduction of replicates of both experimental groups due to the factors aforementioned. Originally, there were three cages, and each cage contains five replicates of each control and treatment groups, but due to these uncontrolled variables, some of the replicates have been contaminated. The researchers disregarded the contaminated units in order to avoid unreliable and inaccurate data.

The significant reduction of the macroalgae from pre- to post-exposure may be attributed to the general herbivory of *Tripneustes gratilla*. Studies suggest that broad-spectrum preference and fast feeding mechanism can be utilized as a potential biocontrol agent, particularly to invasive macroalgae (Stimson et. al., 2007; Conklin and Smith, 2005). Given that the one of the main food preference of *T. gratilla* is a *Padina sp.* and it is also known to feed on *Dictyota sp.*, there is a higher possibility that this is the reason behind the obtained data.

A slight gain of color in some of the corals was visually observed after the exposure to *T. gratilla*. The bleached structures of the corals when pre-exposed to the macroalgae turned to a faint brown color. This suggests that there is a possible recovery from bleaching. However, in order for this statement to be verified, further studies need to be done. The corals which recovered were returned to their natural habitat for them to further recuperate.

Table 2: Computed t-value and p-value between the pre- and post-exposure FMI values

Fleshy Macroalgal Index (FMI)			
Variable	t-value	Df	Significant
Cell count	7.039	14	0.017
Area(cm <sup>2</sup> )	7.114	14	0.014
Coverage percentage	6.787	14	0.023

**Conclusion.** – The reduction of the macroalgal cover may be attributed to the presence of *Tripneustes gratilla*. Although it is only comparably different, the exposure to *Tripneustes gratilla* can potentially aid in the recovery of macroalgal-covered post coral bleached systems and in the control of invasive macroalgae population.

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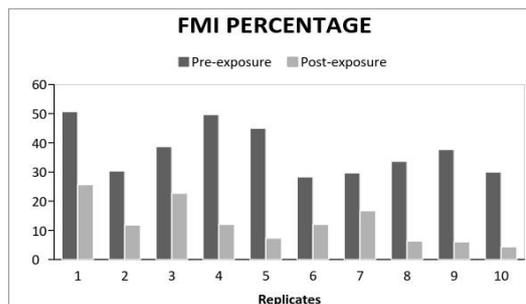


Fig. 3: Comparison of the Fleshy Macroalgal Index percentage values between units before and after exposure to sea urchin

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